

**MONOCHROMATIC FLUID  
TREATMENT SYSTEMS**

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**BACKGROUND**

1. **Technical Field**

The present disclosure relates to systems for photo-processing of fluids, and more particularly to systems for photo-processing of complex fluids, such as blood products, pharmaceuticals, injectable solutions, and vaccines using non-laser light source(s) to generate and deliver monochromatic light at advantageous wavelength(s) in advantageous systems to affect desirable results.

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2. **Background of Related Art**

Efforts have been devoted to treatment regimens for complex fluids, e.g., fluids having medical and/or health-related uses and applications, such as vaccines, pharmaceuticals, injectable solutions, blood products and the like. These efforts have focused in part on technologies and treatment systems to remove, inhibit and/or destroy unwanted fluid components. Treatment regimens aimed at alternative objectives, such as leukoreduction (i.e., the removal of white blood cells to avoid potentially undesirable effects), have also received attention.

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Significant research and development attention has focused on fluids and materials that are collected for transfusion and/or transplantation, e.g., blood products and blood components. In assuring the safety of such materials, significant reliance is placed on pre-transfusion donor evaluation and testing. Despite current efforts, transfusions and transplantations are nonetheless implicated in the transmission of viral, bacterial and protozoan diseases. New infectious agents continue to be identified in the donor

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population, giving rise to increased challenges and concerns among those responsible for ensuring a safe and reliable supply of needed transfusion/transplantation materials.

Testing for unknown pathogens (e.g., HIV in the late 1970's) remains problematic.

Moreover, limitations necessarily exist with respect to the effectiveness of donor

5 screening efforts, e.g., donation volumes, screening/testing impediments, logistical issues, etc. Reliable screening and/or decontamination regimens are of even greater importance for individuals receiving multiple cycles of chemotherapy and/or hematopoietic transplantation because of the cumulative risk associated with repeated transfusion/transplantation episodes.

10 Current research activities and perspectives in the treatment of platelets are discussed in a recently published article. (L. Corash, "Inactivation of Viruses, Bacteria, Protozoa, and Leukocytes in Platelet Concentrates: Current Research Perspectives," Transfusion Medicine Reviews, Vol. 13, No. 1, January, 1999). As noted in the article, risk associated with transfusion-associated infections could be reduced  
15 through development and implementation of decontamination processes that are effective against a broad array of infectious pathogens, regardless of type, including infectious agents not detected through current diagnostic tests. Decontamination processes are preferably effective against cell-free, cell-associated and latent pathogen forms. The need for this breadth in decontamination capability is exemplified by human  
20 immunodeficiency virus (HIV), which is cell-free in plasma, cell-associated in leukocytes, and in latent pro-viral form integrated into genomic leukocyte nucleic acid. Moreover, decontamination processes need to be active against a broad spectrum of

bacteria, including intracellular bacterial forms, to avoid possible bacterial regrowth during storage.

Several potential inactivation technologies for treatment of platelet concentrates have been investigated/described, including psoralens activated with long-wavelength ultraviolet light, merocyanine 540 activated with visible light, riboflavin and methylene blue, and phthalocyanines activated with red light. Of these treatment technologies, attention has focused on the chemistry and associated bonding properties of different compounds, with psoralens receiving the greatest level of attention to date.

Psoralens are planar furocoumarins, many of which are synthesized by plants and ingested as foods. Psoralens preferentially bind to nucleic acid, both RNA and DNA, and vary widely with respect to solubility, nucleic acid affinity, and side reactions, e.g., active oxygen species generation. Investigations have shown that psoralen photochemical treatments can be effective for inactivation of bacteria, RNA viruses and DNA viruses. Exemplary psoralen compounds are described in U.S. Patent No. 5,654,443 to Wollowitz et al.; U.S. Patent No. 5,709,991 to Lin et al.; and U.S. Patent No. 6,171,777 B1 to Cook et al.

However, treatment modalities based on photochemical additives require, by definition, the addition of an external agent/material to the fluid being treated, with the inherent issues and uncertainties associated therewith. Moreover, beyond the requirement that external agent(s)/material(s) be added to the fluid system to effect treatment, other limitations and/or requirements have been identified with respect to certain psoralen-based treatment modalities. For example, long treatment times (up to 4 hours of UVA illumination with 8-methoxypsoralen) and reduced ambient oxygen levels

associated with certain psoralen-based systems are not compatible with requirements for ease of operation, e.g., in clinical blood bank environments. In addition, the addition of a free radical quencher, e.g., rutin (a naturally occurring flavenoid), may be required to prevent platelet damage due to active oxygen species, i.e., to preserve *in vitro* platelet function. The addition of free radical quencher(s) like rutin further increases the complexity of treatment systems.

As discussed by Corash, high UVA doses have also been required for certain psoralen compounds, e.g., from 24 to 70 J/cm<sup>2</sup> for 4'-(amino-methyl)-4,5', 8-trimethylpsoralen (AMT), to treat platelet suspensions in 100% plasma. AMT has also exhibited questionable toxicology profiles. Reductions in plasma protein concentration, e.g., to 15% through use of synthetic platelet additive solution, is effective in reducing energy requirements, e.g., viral inactivation with AMT at 2.4 J/cm<sup>2</sup>, but is not practical for large-scale operations. (Transfusion Medicine Reviews, Vol. 13, No. 1, pp. 21, 23.)

Margolis-Nunno et al. evaluated the treatment of HIV-infected platelet concentrates with an AMT/rutin system using two types of ultraviolet radiation, UVA and UVA1. UVA was characterized as broad-band ultraviolet A light, having a wavelength of between 320 and 400 nm, and UVA1 was characterized as narrow-band UVA light, having a wavelength between 360 and 370 nm. Margolis-Nunno et al. concluded that wavelength was an important consideration in these treatment systems and that, at similar fluences, the tested UVA was more injurious to platelets than was UVA1. (Margolis-Nummo et al., "Psoralen-Mediated Photodecontamination of Platelet Concentrates: Inactivation of Cell-free and Cell-associated Forms of Human

Immunodeficiency Virus and Assessment of Platelet Function In Vivo," Transfusion,  
Vol. 37, September 1997, pp. 889-895.)

Cerus Corporation (Concord, CA) has developed a series of psoralen  
compounds that are being evaluated for their ability to inactivate pathogens in blood  
5 products. In one psoralen-based process under development by Cerus, a platelet  
suspension, which may be pooled from individual units from several donors, is  
transferred to a sterile disposable bag containing Cerus' S-59 psoralen compound. The  
blood-containing bag is illuminated with ultraviolet light for approximately three  
minutes. The Cerus procedure is intended to prevent the pathogen from reproducing and  
10 infecting a transfusion recipient. Cerus envisions the treated platelet suspension being  
ready for transfusion to a recipient.

Photodynamic therapy (PDT) has also received significant research and  
development attention, particularly for cancer indications. In typical PDT treatment  
regimens, a photosensitizer is administered systemically, e.g., a porphyrin derivative, and  
15 after a period in which the photosensitizer accumulates within a target tissue, a measured  
amount of light is applied to the target region. Beyond cancer treatment, PDT treatments  
have been developed for use against ophthalmological conditions, cardiovascular  
conditions (e.g., arteriosclerosis and restenosis), and immune-mediated conditions (e.g.,  
psoriasis).

20 Ultraviolet B (UVB), without the addition of a photosensitizing agent, has  
been shown to inactivate non-enveloped poliovirus in platelet suspensions. However, in  
the reported literature, only small volumes were treated and efficient viral inactivation  
required high UVB doses ( $20 \text{ J/cm}^2$ ) with some decrement in platelet function. UVB

irradiation of full platelet concentrates in larger doses ( $30 \text{ J/cm}^2$ ) has shown efficacy to reduce alloimmune responses to platelet transfusion through leukocyte inactivation.

El-Ghorr and Norval describe the biological effects of narrow-band UVB irradiation, e.g., in treating psoriasis, as compared to conventional broadband UVB irradiation effects. (El-Ghorr et al., "Biological Effects of Narrow-Band (311 nm TL01) UVB Irradiation: A Review," *Journal of Photochemistry and Photobiology B: Biology* 38 (1997), 99-106.) The TL01 lamp tested by El-Ghorr and Norval emits a narrow peak (51% of the radiant energy at 311 nm). Based on limited available test data, the TL01 lamp appears to be more suppressive than broad-band UVB during phototherapy with respect to natural killer cell activity and the function of mononuclear cells, as measured by lymphoproliferation and cytokine production. El-Ghorr and Norval suggest that the noted effect of the TL01 lamp may be both dose related and wavelength dependent.

In a further study, Prodouz et al. evaluated the use of laser-UV in treating blood products to inactivate poliovirus. (Prodouz et al., "Use of Laser-UV for Inactivation of Virus in Blood Products," *Blood*, Vol. 70, No. 2, August, 1987, pp. 589-592.) The Prodouz et al. study uniformly irradiated samples with a XeCl excimer laser that delivered 40 nsec pulses of UV at 308 nm. Prodouz et al. concluded that, when using an excimer laser at 308 nm, there exists a window of efficacy for exposure doses between  $10.8$  and  $21.5 \text{ J/cm}^2$  and peak intensities of less than  $0.17 \text{ MW/cm}^2$  within which a hardy virus is significantly inactivated and platelet and plasma proteins are minimally affected.

Andreu et al. evaluated ultraviolet irradiation of platelet concentrates to reduce HLA immunization by placing suspended platelet concentrates between quartz plates and irradiating with UV-B rays at 310 nm. Andreu et al. concluded that in vitro

function of platelet concentrates remains unaffected by UV-B up to  $2.25 \text{ J/cm}^2$ , but that higher energies inhibit the aggregation induced by ADP and collagen. Andreu et al. identified a gap in treatment effects with UV-B rays between  $0.2 \text{ J/cm}^2$ , where the desired inhibitory effect on immunologic recognition is probably complete, and above  $2 \text{ J/cm}^2$ , where detrimental effect on platelet function appears. (Andreu et al., "Ultraviolet Irradiation of Platelet Concentrates: Feasibility in Transfusion Practice," Transfusion, Vol. 30, No. 5 – 1990, pp. 401-406.)

U.S. Patent Nos. 4,726,949; 4,866,282; and 4,952,812 to Miripol et al. disclose blood product irradiation methods and systems for inactivating white blood cells. The Miripol et al. treatment regimens employ ultraviolet radiation predominantly of a wavelength of 280 to 320 nm, an intensity of  $4 \text{ to } 20 \text{ mW/cm}^2$ , and a total energy exposure of  $800 \text{ to } 20,000 \text{ mJ/cm}^2$ . Eight to twelve conventional, high intensity bulbs are described for use in irradiating blood contained within a flexible, collapsible poly(ethylene-vinyl) acetate plastic bag, the plastic bag typically being stretched on a framework. An exhaust fan is provided in the back of the Miripol et al. apparatus to exhaust heat generated by the high intensity bulbs.

Despite efforts to date, there exists a continuing need for systems that facilitate treatment of complex fluids, e.g., blood products, pharmaceuticals, injectable solutions and vaccines. In particular, systems applicable to high value/complex fluids that effectively and reliably achieve desirable levels of pathogen inactivation, modulation of immune response, medical therapy and/or chemical synthesis, without negatively impacting desirable characteristics of the high value/complex fluid, are needed. While significant attention has been devoted to developing and evaluating a variety of

photosensitive agents in treating pathogens and the like, the potential effects and/or influences of light source(s) and/or wavelength characteristics of light on complex fluid treatment systems have received less intensive study. Indeed, Cook et al. state that treatment of blood products to eliminate transmission of diseases by inactivating pathogens through UV alone is “completely incompatible with maintenance of red cell function.” (U.S. Patent No. 6,171,777 B1, col. 2, lines 58-64.)

### SUMMARY OF THE DISCLOSURE

The present disclosure provides innovative systems for photo-processing of fluids, particularly high value and/or complex fluids, such as blood products, pharmaceuticals, injectable solutions and vaccines. The disclosed systems employ advantageous light source(s) and processing regimens, whether alone or in combination with adjunct additives and/or photoactive agents, to achieve desired results. Preferred systems according to the present disclosure are effective in inactivating pathogens, bacteria and/or viruses, modulation of immune response, and/or leukoreduction, without negatively impacting desirable components and/or attributes of the treated fluid, and achieve desirable results in a broad range of diagnostic, therapeutic and treatment applications. Preferred systems further provide enhanced operating efficiencies and/or processing results in application modalities that employ a broad range of photo-activated and/or photo-responsive materials and/or compounds.

One advantageous aspect of the disclosed systems relates to an ability to utilize light energy to limit and/or minimize degradation of certain fluid properties and/or components, e.g., potential degradation through heat, energy activation or the like, while simultaneously maximizing the desired effect(s) through use of such light energy. The

desired effects may be biologic, e.g., pathogen inactivation, or chemical, e.g., excitation of specific psoralen-based adducts, or a combination thereof. The generation of harmful and/or toxic byproducts is also advantageously minimized or avoided according to the present disclosure.

5                   According to the present disclosure, light source(s) are provided that supply light energy having highly advantageous effect(s) on treated fluids, particularly high value and/or complex fluids such as blood products, pharmaceuticals, injectable solutions and vaccines. The present disclosure further provides system design and processing regimens that, when used in conjunction with disclosed light source(s),  
10                   provide advantageous fluid processing results, both in batch and continuous flow treatment systems.

#### BRIEF DESCRIPTION OF FIGURE(S)

To assist those of skill in the art to which the subject matter of the present disclosure appertains, reference is made to the accompanying drawings and associated  
15                   detailed description, in which:

Figure 1 is a perspective view of one aspect of a treatment apparatus according to an embodiment of the present disclosure;

Figure 2 is a cross sectional view of a first aspect of the treatment apparatus of Fig.1, taken along line 2-2;

20                   Figure 3 is a schematic cross sectional view of a second aspect of a treatment apparatus according to an embodiment of the present disclosure;

Figure 4 is a schematic cross sectional view of combined first and second aspects of a treatment apparatus according to an embodiment of the present disclosure;

Figure 5 is a schematic perspective view of a treatment apparatus according to an alternative embodiment of the present disclosure;

Figure 6 is a schematic cross sectional view of the treatment apparatus of Fig. 5, taken along line 6-6;

5 Figure 7 is a schematic cross sectional view of a treatment apparatus according to a further alternative embodiment of the present disclosure;

Figure 8 is a schematic cross sectional view of a treatment apparatus according to an additional alternative embodiment of the present disclosure; and

10 Figure 9 is a plot of e-coli response based on experimental results obtained according to an embodiment of the present disclosure.

#### DETAILED DESCRIPTION OF PREFERRED EMBODIMENT(S)

The present disclosure provides innovative systems for photo-processing of fluids, particularly high value and/or complex fluids, such as blood products, pharmaceuticals, injectable solutions and vaccines. As used herein, a “complex fluid” is  
15 a fluid that includes a plurality of fluid components that are sensitive to and/or activated by light and/or heat energy, wherein a first light-sensitive/light-activated/heat-sensitive fluid component is to be substantially preserved according to the disclosed treatment regimen, and a second light-sensitive/light-activated fluid component is to be substantially modified, inactivated and/or eliminated according to the treatment regimen.  
20 A “complex fluid” according to the present disclosure may also include added photoactive compound(s)/material(s), e.g., psoralen, and light energy sensitivity/reactivity may be due in part to the presence of such photoactive compound(s)/material(s).

The innovative systems of the present disclosure employ advantageous light source(s) and processing regimens unique to the treatment of complex fluids. The disclosed systems may be employed with or without photoactive compounds and/or agents to achieve desired results, e.g., inactivation of pathogens, bacteria and/or viruses, modulation of immune response and/or leukoreduction. The disclosed systems achieve desirable results in a broad range of diagnostic, therapeutic and treatment applications, and provide enhanced operating efficiencies and/or processing results in application modalities that employ a broad range of photo-activated and/or photo-responsive materials and/or compounds.

According to the present disclosure, light source(s) are provided that supply light energy having highly advantageous effect(s) on treated fluids, particularly high value and/or complex fluids such as blood products, pharmaceuticals, injectable solutions and vaccines. The present disclosure further provides geometric, spectral, intensity and processing regimens that, when used in conjunction with disclosed light source(s), provide advantageous fluid processing results, both in batch and continuous/semi-continuous treatment systems.

Preferred light source(s) according to the present disclosure supply substantially monochromatic light at high average power outputs, but moderate peak power outputs, thereby reducing or eliminating potential damage to high value/complex fluids due to undesirable heating and/or multi-photon initiated processes. Thus, conventional laser light sources and conventional flash lamps are ineffective/less effective, and are to be avoided according to the systems of the present disclosure. Pulsed laser light sources are undesirable and ineffective according to the present

disclosure at least in part due to the high peak power pulses generated thereby.

Conventional high intensity lamps, e.g., mercury lamps, fail to provide the wavelength flexibility, processing efficiency, intensity and carrier media preservation, that are critical to treatment systems according to the present disclosure.

5                   It has been found according to the present disclosure that use of the disclosed high output, monochromatic light sources allows effective exploitation of advantageous treatment windows for high value and/or complex fluids, e.g., blood products, pharmaceuticals, injectable solutions and vaccines. Moreover, use of high output, monochromatic light sources as disclosed herein generate synergistic results in  
10   complex fluid systems that utilize chemical and/or photoactive agents.

                  Before describing specific aspects of preferred treatment systems according to the present disclosure, including attributes of preferred light sources and associated apparatus/treatment regimens, a discussion of contemplated fluid treatment applications and/or techniques in which the disclosed treatment systems offer  
15   advantageous results is provided. This discussion of exemplary fluid treatment applications and/or techniques is provided at a level of detail that will permit persons of ordinary skill in the art to effectively exploit the innovative treatment systems disclosed herein to achieve advantageous results. Additional fluid treatment applications and/or techniques may also be recognized and/or devised for exploitation of the innovative  
20   treatment systems disclosed herein, as will be apparent to persons of ordinary skill in the art from the detailed description that follows, and such additional fluid treatment applications and/or techniques are to be deemed within the scope of the present disclosure.

In a first preferred embodiment of the present disclosure, a system is provided for efficacious treatment of high value/complex fluids that may contain undesirable pathogens, e.g., viral and/or bacterial components, through application of monochromatic light generated by a non-laser light source at advantageous wavelength(s). Exemplary fluids for treatment according to the disclosed treatment regimen include blood products, pharmaceuticals, injectable solutions and vaccines. According to this embodiment, high value/complex fluids are treated to inactivate undesirable pathogens without the addition of chemical additives, photochemical agents and the like, e.g., psoralens, thereby simplifying the treatment regimen and eliminating the need for quenchers and/or post-treatment removal of added materials and/or byproducts.

The disclosed treatment system may be undertaken with respect to high value/complex fluids in real time, i.e., immediately upon the creation and/or availability of the target fluid for treatment. Thus, for example, in the context of blood product or other transfusable/injectable component treatment systems, the disclosed treatment regimen may be advantageously employed immediately upon blood product donation, immediately prior to blood product transfusion, and/or a combination thereof.

Alternatively, treatment systems may be deployed to treat high value/complex fluids immediately prior to aseptic packaging, e.g., in large-scale systems for sterilizing solutions prior to packaging thereof. Each of the foregoing treatment systems may be batch, semi-batch, continuous or semi-continuous in operation.

According to preferred embodiments of the disclosed system, substantially monochromatic light energy is delivered to the desired high value/complex fluid to

achieve maximal absorbance within or by the target pathogen(s), while simultaneously minimizing damage to the non-pathogenic components and/or substrates, e.g., the carrier media, of the treated fluid. Thus, in the case of blood products, it has been found that particularly desirable pathogen inactivation results may be achieved, without deleterious effect to blood product components within the carrier media, by delivering substantially monochromatic light at a wavelength of between 260 and 310 nm, and generally between 270 nm and 308 nm. Preferred monochromatic wavelengths include 282 nm, 290 nm and 308 nm, although additional wavelengths within the preferred ranges disclosed herein are also deemed to be highly desirable. As used herein, substantially monochromatic light generally exhibits a wavelength distribution wherein the light energy falls predominantly within a range of +/- 10 nm relative to the designated wavelength.

In selecting a desired monochromatic light wavelength, it has been found that wavelengths below about 260 nm are effective in inactivating pathogens within blood systems, but have a significantly increased tendency to damage blood components that are to be safeguarded. Conversely, it has been found that wavelengths above about 315 nm have a markedly reduced tendency to achieve desired/required levels of pathogen inactivation. Pulsed laser light sources have also been found to affect an unacceptable level of damage to the carrier media.

In a further embodiment of the present disclosure, a treatment system is provided in which high value/complex fluids may be treated to reliably and efficaciously inactivate pathogen(s) contained therewithin, wherein the treatment system includes one or more photoactivated compounds and/or additives. Thus, according to the disclosed treatment system, conventional photoactivated compound(s) and/or additive(s) are

combined with a treatment fluid, e.g., a blood product, pharmaceutical, injectable solution and/or vaccine, as is known in the art. However, unlike prior art treatment regimens, the disclosed system advantageously irradiates the combined fluid, i.e., high value/complex fluid and photoactivated compound/additive combination, with

5 substantially monochromatic light energy from a non-laser light source at preferred wavelength(s) and intensity(ies), and in preferred geometric/structural arrangement(s), as described in greater detail hereinbelow.

According to the disclosed treatment regimen that includes a conventional photoactive compound/additive, e.g., one or more psoralens, dimethylmethylene blue,

10 riboflavin and the like, pathogen inactivation may be enhanced while minimizing unwanted side effects by irradiating such fluid systems with substantially monochromatic light at wavelength(s) between 270 and 340 nm. Particularly preferred wavelengths include 282 nm, 290 nm, 308 nm and 320 nm. Light delivery to the treatment fluid is controlled such that the wavelength is adjusted and peak intensity levels minimized,

15 thereby advantageously exciting the photoactive compound(s)/additive(s) to achieve the desired results. Thus, in psoralen-based embodiments of the present disclosure, sufficient light intensity is delivered within a narrow spectral range to excite specific adducts.

According to such treatment regimens, desirable results are achieved, including inhibiting target pathogen(s) from reproducing/infecting a transfusion recipient, and/or generating

20 improved immune response. Moreover, the systems of the present disclosure provide dose rates that speed these processes.

It is further contemplated according to the present disclosure that particularly advantageous results may be achieved by illuminating/irradiating a fluid

system, e.g., a blood system that includes a photoactive compound/additive, so as to achieve the desired activation of the photoactive compound/additive and to additionally effect dissociation of the compound/additive. It is contemplated that the dual processing objectives may be achieved as part of the inactivation process, or in a subsequent monochromatic light-based treatment, e.g., in a continuous processing treatment system. Generation of an optimally effective adduct, e.g., an adduct that is wavelength dependent, or generation of an optimal surface structure that is wavelength dependent facilitates successfully accomplishing both desired results according to the disclosed treatment system.

10 In another preferred embodiment of the present disclosure, monochromatic light from a non-laser light source is advantageously utilized to effect leukoreduction to minimize or prevent blood platelet alloimmunization, i.e., the transfer of pathogenic immune response information through blood transfusion. Thus, according to a preferred embodiment of the present disclosure, a blood system is treated with monochromatic light having a wavelength at between 260 and 310 nm, generally between 270 and 308 nm, and preferably at a wavelength of 282, 290 or 308 nm. A preferred blood system is treated at an intensity of greater than 20 mW/cm<sup>2</sup> to effect desired rates of leukoreduction. Substantially monochromatic light energy having the desired UV-B wavelength is typically generated from a non-laser light source, thereby moderating energy levels in the manner desired according to the disclosed treatment regimen.

It is further contemplated according to the present disclosure that monochromatic light generated by a non-laser light source at predetermined wavelengths will exhibit synergistic effects with different adducts in photophoresis treatment

regimens. Thus, it is contemplated that monochromatic light may be used in connection with extracorporeal photophoresis, AIDs treatment, cancer treatment (e.g., cutaneous T cell lymphoma (CTCL)), Lupus treatment, and other disease treatments and/or treatment regimens directed to conditions requiring modulation of the immune system.

5                    Additionally, it is contemplated that graft vs. host disease (GVHD) may be treated through systems utilizing monochromatic light of a predetermined wavelength and intensity. GVHD treatment regimens may be achieved that optimally inactivate T-cells while preserving other substrates and components to enhance the likelihood of transplant success. In particular, with stem cell transplants, photoactivation with an  
10                    optimal light energy is believed to maximize inactivation of T-lymphocytes and minimize damage to the hematopoietic stem cells.

                    Thus, for example, treatment regimens involving photoactive compounds and/or agents having efficacy in treating the above-noted diseases and/or conditions are believed to exhibit advantageous treatment results when activated with monochromatic  
15                    light, particularly monochromatic light generated by a non-laser light source, at wavelengths of between 250 and 400 nm, and preferably 260 to 310 nm.

                    Additional treatment regimens utilizing substantially monochromatic light are also contemplated according to the present disclosure, including body irradiation techniques and treatment of a variety of medical and health-related fluids. Thus, in a  
20                    preferred embodiment of the present disclosure, monochromatic light delivery systems are provided that accommodate irradiation of one or more portions of a body, e.g., a limb, extremity or the like. The light delivery system may advantageously permit introduction of the body portion to an enclosed chamber and/or entail paddle light sources that are

easily and reliably positioned adjacent the body region to be treated and/or provide faster, more comfortable systems for whole body irradiation. Upon irradiation of the target body region with monochromatic light of a desired wavelength and intensity, photoactivation of the surface layer(s) thereof may be achieved with desirable result.

5                   It is additionally contemplated according to the present disclosure that monochromatic light of predetermined wavelength and intensity may be used to advantageously and reliably sterilize fluids, e.g., saline, interperitoneal fluid, vaccines, liquid pharmaceuticals, injectable fluids, and other sterilizable fluids. In particularly preferred embodiments, medical fluids are processed into a relatively thin film and  
10   exposed to substantially monochromatic light generated by a non-laser light source at a wavelength between 260 nm and 310 nm to achieve advantageous, uniform dosing. Alternatively, thicker flow streams may be processed, with engineered turbulence to achieve desired uniformity in dosing. Thus, improved sterilization results may be achieved for a range of fluids according to the treatment systems disclosed herein.

15                   It is further contemplated according to the present disclosure that monochromatic light generated by a non-laser light source at predetermined wavelength(s) and intensity(ies) may be utilized to activate/promote chemical synthesis and/or chemical processing. Thus, the activation energy associated with a specific process step and/or the kinetic parameters associated with a particular chemical process  
20   may be supplied by such monochromatic light, thereby potentially boosting yields and speeding process rates. For example, processes that are biologic in nature, e.g., activation/inactivation of biologic organisms influencing chemical processes, or chemical in nature, e.g., dependent upon the excitation of specific active site(s) and/or functional

group(s), may be promoted and/or enhanced through monochromatic light energy supplied according to the present disclosure.

In all cases, dose determination is not straightforward in opaque fluids. Previous disclosures have often mistaken surface dose with dose delivered to the fluid volume. The large area sources disclosed herein allow dose to be advantageously controlled through changes/modifications in light source geometry. Typical systems require severe overdosing at some volumes to ensure adequate dosing of others. The disclosed treatment systems, however, provide a means to treat complex, opaque fluid volumes more uniformly.

In short, substantially monochromatic light having predetermined wavelength, geometry and intensity characteristics may be exploited to achieve and/or deliver significant benefits in a wide range of applications, systems and techniques according to the present disclosure. Through control of the wavelength, intensity and delivery characteristics of the monochromatic light, advantageous results are achieved in a plurality of fluid applications, with a high degree of specificity and in an economically desirable manner. Having thus described a series of applications and techniques that may advantageously exploit monochromatic light of predetermined wavelength, attention is now turned to preferred systems for generating and delivering monochromatic light to treatment fluids according to the present disclosure.

With reference to Figures 1 and 2, a treatment apparatus 100 is depicted for advantageously treating high value/complex fluids according to the present disclosure. Treatment apparatus 100 is configured and dimensioned to receive an advantageous light source for generating substantially monochromatic light according to

the present disclosure, and is configured to treat high value/complex fluids positioned external thereto, e.g., blood products, in batch or semi-batch treatment regimens.

Treatment apparatus 100 is merely exemplary of preferred treatment apparatus according to the present disclosure, as will be apparent from the discussion that follows.

5                   Treatment apparatus 100 includes a housing 102, an inlet port 104 adjacent a first end of housing 102, and an outlet port 106 adjacent an opposite end of housing 102. Housing 102 includes an outer wall 103 that defines a substantially square cross section. End flanges 107a, 107b are provided at either end, each end flange 107a, 107b defining a substantially circular surface 109 that bounds an opening 110. Flanges 10   107a, 107b are typically welded to outer wall 103 to define a fluid-tight volume 105 (with an inserted light source, as described below) which is in fluid communication with inlet and outlet ports 104, 106.

Outer wall 103 and the inner faces of end flanges 107a, 107b may be advantageously fabricated from, or surface treated with, a reflective material, e.g., 15   aluminum. Such reflectivity assists in directing ultraviolet radiation toward high value/complex fluids being treated by apparatus 100, as described hereinbelow. Outer wall 103 and end flanges 107a, 107b may be fabricated from a relatively rigid material having sufficient strength to withstand the fluid pressure and fluid flow within volume 105. Aluminum is a preferred material for use in fabricating outer wall 103 and end 20   flanges 107a, 107b, although other metals are also contemplated.

In a preferred embodiment of the present disclosure, housing 102 functions as a ground for a light source positioned within volume 105. In such instance, outer wall 103 of housing 102 is necessarily fabricated from a conductive material, e.g.,

aluminum, and is electrically grounded, as is known in the art. The importance of the grounding properties of housing 102 to advantageous operation of preferred systems according to the present disclosure will become more readily apparent from the light source discussions which follow.

5                   Inlet and outlet ports 104, 106 advantageously communicate with a source of cooling fluid (not pictured), preferably a cooling fluid that is substantially transparent to ultraviolet radiation. For example, inlet port 104 may be connected to a source of cooling water that advantageously flows through volume 105, and exits housing 102 through outlet port 106. Cooling water may be advantageously recycled to housing 102  
10 and/or passed through a light source (either before or after passage through volume 105), as described in greater detail hereinbelow. The flow rate of the cooling fluid within volume 105 is controlled through conventional means, e.g., valving, available water pressure and/or pump settings.

                  A quartz plate 108 is centrally mounted on one face of housing 102 by  
15 mounting flange 112 to facilitate passage of light energy from a light source (not pictured in Figs. 1 and 2) positioned within circular passage 110 to high value/complex fluids positioned external thereto. Quartz plate 108 is advantageously defined by three individual quartz panels 108a, 108b, 108c to provide a desired level of light energy transmission from housing 102 for treatment of high value/complex fluids positioned  
20 adjacent quartz plate 108. Cross beams 112a that separate quartz panels 108a, 108b, 108c from one another advantageously provide additional grounding to a light source positioned within housing 102, as described below. The external surfaces of quartz panels 108a, 108b, 108c define treatment surfaces upon which, or in substantial

juxtaposition with which, high value/complex fluids may be treated according to the present disclosure. By placing the high value/complex fluid in direct contact with quartz plate 108, minimal reflection of the monochromatic light delivered through cooling fluid-containing volume 105 and quartz panels 108a, 108b, 108c is permitted.

5                   Of note, high value/complex fluids, e.g., blood products, may be positioned on the external surfaces of quartz panels 108a, 108b, 108c within storage vessel(s) or container(s), e.g., treatment bag(s), that are highly transparent to ultraviolet radiation, e.g., flexible bags fabricated from poly(ethylenevinyl acetate), as are known in the art. The storage vessel/container is preferably free of leachable materials. The high  
10   value/complex fluids within the storage vessels/containers are advantageously substantially flattened to define a relatively uniform fluid depth or thickness to be treated according to the present disclosure. Mixing of the complex fluid may be provided through appropriate mechanical means, if desired, during irradiation treatment.

                  According to preferred treatment systems according to the present  
15   disclosure, a non-laser light source is introduced through opening 110 in flanges 107a, 107b for treatment of high value/complex fluids positioned adjacent or in juxtaposition with quartz panels 108a, 108b, 108c.

                  With reference to Figure 3, exemplary non-laser light source 200 is schematically depicted in cross section. Light source 200 is substantially cylindrical in  
20   geometry and is advantageously configured and dimensioned for introduction into opening 110 of housing 102. Light source 200 advantageously rests on circular surfaces 109, thereby centering light source 200 within housing 102 and defining a fluid-tight volume 105. The ability to introduce and withdraw light source(s) from housing 102 is

clearly advantageous in numerous respects, including an enhanced ability to service light source(s), as needed, and increased flexibility in utilizing alternative light source(s) within housing 102, e.g., to generate substantially monochromatic light energy of varying wavelengths. Alternatively, a light source may be integrally associated with housing 102, i.e., non-removable with respect thereto. In such case, it is contemplated that the photon-producing gas(es) contained within the light source may be replaceable/substitutable. Thus, in either case, apparatus 100 provides significant versatility in the treatment of high value/complex fluids with light energy of varying wavelengths.

Monochromatic light source 200 includes an outer wall 202 and an inner wall 204 that cooperate to define an annulus 206. An elongated conductor 208 extends the length of light source 200, preferably along the central axis of the light source's cylindrical geometry. A substantially cylindrical region 210 is defined between inner wall 204 and elongated conductor 208. Cylindrical region 210 advantageously communicates with inlet and outlet ports for ingress and egress of cooling fluid, e.g., cooling water. End flanges are provided to define fluid tight seals for annulus 206 and cylindrical region 210. Thus, monochromatic light source 200 generally includes three elongated, concentric elements: elongated conductor 208, cylindrical region 210 and annulus 206.

According to the present disclosure, light emitting, i.e., photon-producing, gas sources are contained within the bounded volume of annulus 206. Preferred light emitting/photon-producing gas sources are "excimers," as are known in the art. Excimer sources (also called "excited dimers") driven by dielectric barrier discharge offer a unique, advantageous set of capabilities for fluid treatment regimens according to the

present disclosure: operation at ambient temperature, discretely tunable monochromatic output, variable emitting areas, and high germicidal UV output per lamp. Dielectric barrier discharge technology is known for its ability to produce energetic molecular species at ambient temperature discharge and is also used to generate ozone. Applicable voltages, frequencies and currents are known to those of ordinary skill in the art. This excitation source allows even high power excimer lamps to operate at the same temperature as the fluid to be treated based on the advantageous geometric/structural arrangements disclosed herein. As described herein, excimer sources may also be built in a variety of advantageous geometric configurations.

10                    Monochromatic light source 200 emits light uniformly over its entire surface area and is configured to operate at reduced temperatures, particularly based on the system designs disclosed herein. The substantially monochromatic output of light source 200 can be tuned to produce high spectral irradiance (watts/nm) within peaks of the process action spectra to maximize the effectiveness of the light energy, e.g., for pathogen inactivation, leukoreduction, and the like. Excimer sources advantageously produce light output wherein 90% of the output is within a 10 nm band, and preferably a 5 nm bandwidth. The light output may be discretely adjusted across VUV, UV-A, UV-B and UV-C wavelengths by changing the rare and/or halogen gases used. Efficiencies vary with gas mix and geometry from 10 % to greater than 30% efficiency, with demonstrated input powers from less than one watt to greater than 10 kW.

                    According to the present disclosure, it has been found that monochromatic light emission is required to optimally process most fluids due to the wavelength-specific absorbance characteristics of the fluid, the preservation of critical fluid functions, and/or

the efficient delivery of light to produce a desired effect. An irradiation device where the emitting surface of the irradiator is both temperature controlled and in contact with the target is highly desirable. In this way, the unit efficiently delivers light, and controls the temperature of the sample. This can also be viewed as monochromatic emission since  
5 heat energy is infrared radiation. For example, while the use of UV-B for treating blood is known, an optimal system would produce only UV-B. Shorter wavelengths, it is understood, damage the blood component. Less appreciated is that longer wavelengths can result in repair of inactivated organisms in a disinfection process, unintended byproduct formation when chemical additives are used and, in all cases, heating by  
10 infrared emissions.

Higher intensity sources generally result in faster processing times.

However, very intense light sources, like lasers and flash lamps, can also generate non-linear effects including surface ablation and "two-photon excitation." Therefore, for all first order processes, defined here as processes that are dependent on dose and/or the  
15 number of absorbed photons, a continuous-duty light source as disclosed herein provides optimal dose rates, while minimizing unwanted effects due to high intensity. High average power is therefore preferred because it speeds process times.

According to the present disclosure, excimer light sources may be provided to deliver a variety of wavelengths based on the gas(es) utilized. Preferred  
20 excimer light sources/gas(es) are summarized in Table 1.

Table 1

	<u>Excimer Gas(es)</u>	<u>Wavelength</u>
	XeI	253
	Cl <sub>2</sub>	259
5	XeBr	282
	Br <sub>2</sub>	289
	XeCl	308
	Filtered XeBr	320
	I <sub>2</sub>	342
10	XeF	351

With reference to Figure 4, light source 200 is typically positioned within apparatus 100 by sliding light source 200 through opening 110. Cooling fluid is typically supplied to cylindrical region 210, e.g., cooling water from a conventional source. Of  
15 note, cooling fluid may be supplied in series to apparatus 100 and light source 200. Such cooling fluid may be supplied first to apparatus 100 or to light source 200, and may flow co-currently or countercurrently within the respective fluid-tight regions. Treatment fluid 250 is bounded by vessel 252, e.g., a blood bag, and positioned adjacent to or in  
juxtaposition with quartz panels 108a, 108b, 108c within flange 112, i.e., external to  
20 housing 102. In a preferred embodiment of the present disclosure, the treatment fluid is a blood product, pharmaceutical, injectable solution or a vaccine.

Light source 200 is energized to deliver monochromatic light energy by supplying alternating voltage to elongated conductor 208 and establishing housing 102 as a ground for the overall system. Voltage supplied to elongated conductor 208 typically  
25 falls within a range of 100 to 10,000 volts. The capacitive coupling between the high voltage within elongated conductor 208 and grounded housing 102 excites the photon-producing excimer gas(es) contained within the bounded volume of annulus 206. Based on the excimer gas(es) contained within annulus 206, substantially monochromatic light

of a substantially uniform wavelength is generated and delivered from annulus 206, i.e., across substantially the entire surface area of outer wall 202 and inner wall 204. The monochromatic light energy is advantageously transmitted through the cooling fluid contained within volume 105 and quartz plate 108 to treat the treatment fluid 250 within vessel 252 that is positioned adjacent to or in juxtaposition with quartz panel 108. The internal reflectivity of housing 102, if any, contributes to maximizing the amount of monochromatic light that is transmitted through quartz plate 108 for treatment of treatment fluid 250. In addition, reflector(s) may be added to increase flux through quartz plate 108.

10 In preferred embodiments of the present disclosure, light source 200 contains excimer gas(es) selected from those set forth in Table 1 hereinabove. Treatment fluid 250 is advantageously selected from among blood products, pharmaceuticals, injectable solutions and vaccines, although it is further contemplated that other fluids may be treated according to the present disclosure, as described hereinabove. It is further contemplated that treatment fluids positioned adjacent to, or in juxtaposition with, quartz plate 108 may contain photoactive compounds and/or materials, e.g., psoralens and the like, that are activated or otherwise affected by the monochromatic light delivered by light source 200.

Preferred treatment parameters vary depending on several factors, including the characteristics of the fluid being treated, the desired outcome of the treatment, and the wavelength of the monochromatic light being transmitted by light source 200. However, it is generally contemplated that fluid treatments according to the present disclosure will involve delivery of monochromatic light having a wavelength of

between 250 and 400 nm; for pathogen inactivation preferably between 260 and 310 nm; and for chemical excitation preferably from 220 to 350 nm. The monochromatic light treatment typically involves surface dosages of 0.1 to 10 J/cm<sup>2</sup>, based upon intensities of 2 to 50 mW/cm<sup>2</sup> and treatment times of 5 seconds to 15 minutes.

5                Several unique advantages are apparent from the fluid treatment regimens described herein. Given the susceptibility of treatment fluids to undesirable heating during treatment, the advantageous continuous cooling of the treatment fluids by cooling fluid within volume 105 prevents deleterious effects to the treatment fluids during treatment with monochromatic light according to the present disclosure. Beyond cooling,  
10        “cooling fluid” that flows through volume 105 may be used to control the temperature of the treatment fluid at any desired temperature level, depending on the requirements of the treatment system. Thus, it is contemplated that the “cooling fluid” may be used to heat and/or moderate the temperature of the treatment fluid, as desired by users of the disclosed system. Moreover, cooling fluid flowing within cylindrical region 210  
15        advantageously facilitates efficient cooling of light source 200.

              Thus, unique aspects of the monochromatic light generation system according to the present disclosure, wherein an excimer light source is positioned within a fluid-tight, cooling fluid volume, permit reliable and efficacious cooling (or temperature moderation/control) of a treatment fluid through direct heat transfer between  
20        the treatment fluid and a substantial heat sink in close proximity thereto. Indeed, according to the present disclosure, it is possible to ensure negligible heat gains and/or temperature change within a treatment fluid throughout the duration of a treatment

regimen. The transmission characteristics of water permit operation of this design with a wide range of wavelengths.

In addition, monochromatic light energy within the desired UV wavelength ranges that is delivered to treatment fluids according to the present disclosure is advantageously generated and transmitted across a substantial surface area, the geometry of which advantageously conforms or corresponds to the surface area geometry of the treatment fluid. Thus, unlike conventional high intensity bulbs and/or elongated fluorescent light generation apparatus, which generate light from a "point" or "linear" source, the disclosed treatment system advantageously generates monochromatic light through a surface. The shape of this light emitting surface is directly associated with the fluid-tight enclosure/housing, e.g., mounted thereto. In the disclosed embodiment of Figs. 1-4, the light emitting surface geometry is defined by quartz panel 108 and is substantially planar. The geometry of the treatment surface of the treatment fluid substantially conforms or corresponds to such planar emitting surface, i.e., the treatment surface geometry is also substantially planar. The advantageous relationship between the light emitting surface geometry and the fluid treatment surface geometry translates into reliable and efficacious energy delivery to the treatment fluid.

Thus, according to the present disclosure, light source(s) for fluid or surface treatment are advantageously provided wherein the surface area geometry of light source emission conforms/corresponds to the surface area geometry of the fluid and/or fluid container to be treated. Surface emitter sources that emit uniformly across a large area can be placed close to the target - providing a very small footprint, high intensities, and a uniform irradiation field. While a planar arrangement as depicted in Figs. 1-4

provides efficient energy delivery to the treatment fluid, it is contemplated that the fluid container may be a tube, where the light source is an annulus irradiating inwardly. The container may also be a thin sheet, in which case the light source is a plane. The container may be a bag, in which case the light source may be a hollow volume, approximating the shape of the bag (similar to a mold) which radiates inwardly. The container may also be an annulus, in which case the light source may be a cylinder radiating outwardly, or an annulus (of large diameter) radiating inwardly. In all cases, the surface area of the light source is engineered to match the fluid container, providing a more uniform dose distribution to the treated volume. For photodynamic therapies (PDT), the concept of "vessel" or container for the treatment fluid can be extended to include the human body or a limb of the body, for whole body irradiation or skin treatment. Hence, light sources that are surface emitters, as disclosed herein, rather than point sources or linear sources, offer significant treatment advantages for high value/complex fluids, as described herein.

While preferred systems according to the present disclosure have been described in some detail with reference to the embodiments of Figs. 1-4, a range of alternative structural and geometric arrangements for delivering substantially monochromatic light energy are contemplated. For example, as shown schematically in Figures 5 and 6, apparatus 300 may be utilized to generate and deliver monochromatic light from a light source 350. Light source 350 includes an annulus 360 within which excimer gas is contained. Alternating voltage is supplied to a conductive screen 370 positioned adjacent an inner wall 365 of annulus 360, and a housing (not pictured) that may advantageously surround light source 350 typically provides a ground with respect

thereto. Housing 350 may also function as a reflector to direct light energy inward. A fluid-tight annulus 330 is provided interior of excimer gas-containing annulus 360. A cooling fluid typically flows within fluid-tight annulus, e.g., cooling water or air.

Treatment fluid 380 to be treated by apparatus 300 typically flows through central tubular region 390. Tubular region 390 may be an integral component of apparatus 300, or may be defined by tubing that is introduced to and/or threaded through along the central axis of apparatus 300, e.g., as blood is obtained from or transfused to an individual. Tubing may be advantageously fabricated from Teflon or the like.

Fluid to be treated by apparatus 300 may be selected from among blood products, pharmaceuticals, injectable solutions, vaccines and other fluid systems.

Cooling fluid within cooling annulus 330 advantageously provides immediate cooling (or temperature control/moderation) of treatment fluid 380 passing through tubular region 390, thereby ensuring that undesirable temperature changes do not damage or negatively impact the treatment fluid 380. Moreover, monochromatic light is generated and inwardly transmitted toward treatment fluid 380 across a substantial surface area, i.e., the surface area of inner wall 365. The light emitting surface area geometry across which monochromatic light is transmitted to treatment fluid 380 conforms or corresponds to the treatment surface geometry for treatment fluid 380 as it passes through central tubular region 390, i.e., an annular surface geometry. This system design (Figs. 5 and 6) has particular advantages for transfused blood products where handling of the fluid is to be minimized and tubing is already used to transport fluid from donor to storage to recipient.

With reference to Figure 7, a cross section side view of a further alternative treatment system according to the present disclosure is depicted. Apparatus

700 includes a symmetric pair of photon-emitting excimer light sources 702a, 702b, each enclosed within a housing defined by an inner wall 704 and an outer wall 706. The external walls 708 of apparatus 700 are grounded at grounds 710. A screen 712 is provided adjacent to, but internal of, inner wall 704. A treatment fluid 720 is contained within a vessel 722. A fluid-tight region 724 separates vessel 722 from inner wall 704 of light sources 702a, 702b. A cooling fluid and/or gas is generally located within region 724. The cooling fluid/gas may be statically contained within region 724 or may flow therethrough, to provide a desired level of temperature control/moderation. This design has particular applicability to extracorporeal photophoresis systems.

In use, alternating voltage is delivered to screen 712. The voltage generated between screen 712 and ground 710 excites the excimer gas(es) contained within light sources 702a, 702b. Alternating voltage may be applied between grounds 710a and 710b to excite both light sources 702a, 702b from a single power supply. Monochromatic light is generated and delivered through inner wall 704, the cooling fluid/gas contained within fluid-tight region 724, and vessel 722 so as to treat fluid 720 contained therewithin. The light emitting surface geometry conforms/ corresponds to the treatment geometry, i.e., both are substantially planar. Moreover, cooling fluid/gas within region 724 provides advantageous temperature control/moderation to treatment fluid 720 throughout the treatment regimen. Of note, apparatus 700 may be of a relatively large scale and may operate as light emitting paddles or as radiating walls for an enclosure for use in whole body and/or limb monochromatic light treatments.

With reference to Figure 8, apparatus 800 is schematically depicted according to the present disclosure. Apparatus 800 includes a substantially elliptical

annulus 802 that contains excimer gases for monochromatic light generation and emission. A substantially elliptical high voltage screen 804 is positioned within annulus 802 within a fluid-tight region 806. A treatment fluid 808 is positionally centrally to apparatus 800 within a container 810 that defines a substantially elliptical outer geometry. Container 810 may be a blood bag or the like. The outer housing of apparatus 800 is grounded (not pictured) to allow excitation of the excimer gases within annulus 802 upon delivery of high voltage to screen 804. Fluid-tight region 806 advantageously contains a cooling fluid/gas to control/moderate the temperature of treatment fluid 808 throughout the treatment thereof. The light emission surface area geometry (substantially elliptical) advantageously conforms to the treatment surface defined by container 810 (substantially elliptical).

As is readily apparent from the preceding description, an elliptical design is merely exemplary of a non-symmetric geometric design contemplated according to the present disclosure. However, such elliptical design is not to be deemed limiting of the scope of the present disclosure, and it is to be understood that other non-symmetric geometric designs are contemplated and offer potential advantages to various treatment regimens.

As with most treatment fluids, in treating blood products the light source must be configured to operate reliably and economically. Blood products are opaque, i.e., they absorb and scatter light over small distances. Blood is also a delicate fluid requiring temperature control and gentle mechanical handling. Blood bags are optimized for the safe and stable storage of blood products, for optimal handling, and to minimize potential contamination, not for efficient irradiation. Based on inherent limitations

associated with blood bags and other blood containers, it would be highly desirable to treat blood products before they enter storage bags in the first instance or upon egress therefrom. In other words, it is highly desirable to provide an irradiation device for blood products that accommodates real time treatment of flowing blood.

5                   A preferred embodiment for providing real time treatment of flowing blood according to the present disclosure utilizes a light source that provides a targeted dose to blood flowing in a tube, e.g., utilizing apparatus 300 of Figs. 5 and 6, to blood flowing in a sheet or plane, e.g., as shown in Fig. 7, or similar system designs. Several design points are used to optimize preferred systems according to the present disclosure.

10   First, as with any opaque fluid, light intensity varies greatly over small distances as light is absorbed (and scattered) by the fluid. Delivery of a uniform dose usually requires thin films. The use of optimized light emitting surfaces will have advantages for treating cylindrical flows that minimize the dose distribution that complicates treatment of thin films, including the balancing effects of absorbance and converging light.

15                   While light intensity drops off with distance as a function of absorbance, following Beer's law, dose is dependent on intensity. Therefore, dose is dependent on the thickness of the fluid sample. Over-exposure of some target material may be required to adequately dose another part of the target. Since turbulence can damage blood products, opportunities to use mixing are generally limited. Thin films have therefore

20   been used in an effort to deliver a more uniform dose. In addition to geometric issues, dose is time dependent. Flow can be optimized to minimize dose range. This is not easily achieved with blood products.

In this regard and with reference to apparatus 300 depicted in Figs. 5 and 6, it is noteworthy that central tubular region 390 may receive treatment fluid directly, or in an alternative embodiment, central tubular region 390 may receive a tube or pipe threaded therethrough. Thus, for example, in blood donation and/or blood transfusion systems, tubing passing to or from a patient's body may pass through tubular region 390, thereby permitting the blood stream to be treated with monochromatic light from light source 350 in real time, i.e., in the process of blood donation and/or in the process of blood transfusion to a recipient. Such blood streams receive immediate and continuous cooling from cooling fluid that passes through cooling annulus 330.

Inasmuch as annulus 360 (which contains the excimer gas) surrounds tubular region 390 in apparatus 300 of Figs. 5 and 6, light source 350 advantageously simultaneously delivers monochromatic light to a treatment fluid within tubular region 390 from all directions. The structural and geometric arrangement thus optimizes light intensity distribution to the treated fluid, by transmitting light energy inwardly from an annular light transmission surface. The flow of the treatment fluid through tubular region 390 may also be substantially non-turbulent, thereby minimizing the potential damage to treatment fluids passing therethrough.

Transfer tubes thus offer an advantageous geometric opportunity because light transmission reaches the treatment fluid from multiple directions simultaneously. A tube irradiated from all sides can have a near optimal uniform dose distribution. Since the proposed light source delivers a uniform intensity to the entire surface area of the tube. While light is absorbed as it travels toward the center of the tube, this effect is opposed by the smaller effective area the light flux must pass through. Total

optimization may require only an adjustment of the tube diameter, as is well within the skill of persons skilled in the art.

Beyond the inward irradiation systems disclosed in Figs. 5-8, it is further contemplated that the fluid flow may be through an annulus surrounding the excimer gas.

5 In this alternative embodiment, monochromatic light would be radiating both outwardly and inwardly through the treatment fluid, with many of the benefits of apparatus 300 disclosed hereinabove, e.g., real time treatment, reliable cooling, etc. Of note, in apparatus 300 and other flow designs according to the present disclosure, it is contemplated that treated fluid may be recycled (in whole or in part) and/or fed to further  
10 treatment apparatus aligned in series for further processing, as may be needed to achieve the desired treatment results. Indeed, it is contemplated that a series of treatment apparatus may be arranged in series, with multiple light sources introducing light energies of differing intensities and/or wavelengths to respective treatment apparatus, thereby expanding the treatment regimen to target different pathogens and the like.

15 Thus, treatment apparatus according to the present disclosure are not limited to the exemplary designs shown in the accompanying figures, but may take a variety of geometric forms and structural arrangements, as will be apparent to persons skilled in the art based on the detailed description provided herein. Alternative treatment apparatus may thus be utilized, provided the desired monochromatic light irradiates the  
20 treatment fluid for a time and to a degree necessary to achieve the desired results, without imparting undesirable heating of the treatment fluid.

To further illustrate advantageous applications of the disclosed treatment systems, several examples are provided that illustrate certain specific aspects of the

present disclosure. However, it is to be understood that these examples are merely exemplary of applications, techniques, systems, methods and apparatus utilizing the disclosed technology, and are not to be limitative thereof. Many variations and alternative applications of the disclosed technology that fall within the spirit and scope of the present disclosure are contemplated and will be apparent to those of skill in the art from the detailed description provided herein, and the examples that follow.

#### EXAMPLE 1

As depicted in Figs. 1-4, apparatus 100 was utilized with light source 200 positioned therewithin to simulate processing of blood products using a photochemically activated process. XeBr was selected as the excimer gas. The output was filtered using window glass to remove 282 nm light and pass only the emission centered at 320 nm. Blood products contained within conventional blood bags were positioned atop quartz plate 108 and irradiated with monochromatic light from the XeBr excimer light source. Cooling fluid flowing through the treatment apparatus maintained the blood samples at a constant temperature, and prevented any deleterious effect thereto. Platelets were maintained at 24°C, based on the cooling fluid being supplied to the system at that temperature and the platelet bag being in intimate contact with the relatively large surface area of the quartz panels.

Based on irradiations with the XeBr excimer source using the apparatus of Figs. 1-4, no clinically significant effect on platelets was observed for exposures greater than 4 J/cm<sup>2</sup> measured as a change in platelet count (immediately after irradiation or 24 hours later), CD62, morphology score, LDH, pH, pO<sub>2</sub>, PCO<sub>2</sub> or bicarbonate, osmatoc recovery. Similarly for plasma irradiations > 4J/cm<sup>2</sup> showed no effect on PT< PTT,

fibrogen or Factor VIII. For red cells, measurements included hematocrit, LDFH, 2,3 D-P-G, % hemolysis and others. No changes were observed with surface doses greater than 4 J/cm<sup>2</sup>.

Based on these test results, it was concluded that the disclosed treatment system was effective in treating blood product samples without causing deleterious effects thereto.

## EXAMPLE 2

These experimental studies were designed to show that refractory viral contaminants can be inactivated using 282 nm light in real blood products using the treatment systems of the present disclosure. The system was evaluated for its ability to inactivate porcine parvovirus (PPV). PPV (NADL-2 strain) is an 18 - 26 nm, non-enveloped, DNA-containing parvovirus, which exhibits a high degree of resistance to a range of physico-chemical reagents. It is not treated using cell washing techniques because it is non-enveloped. PPV inactivation using psoralen and riboflavin are inefficient because the small genome size of the virus requires either larger doses of activating compound or longer irradiation times.

The strength (titer) of the production lot of virus used in the study was determined by a plaque assay utilizing ST indicator cells. GLP lab standards were maintained. The PPV stock solution used in this example tested positive for identity when tested with a polyclonal antisera specific for PPV and is free of potential bovine (BAV, BPI3, BPV, BVDV, IBR, and REO-3) and porcine (PAV and TGE) viral contaminants.

The blood components were not diluted, and were treated as they would be stored in a blood bank. Volumes used (30 to 150 ml) and the size of bags simulated the thickness of blood product in standard storage bags. Samples which altered the virus titers by  $>0.5 \log_{10}$  were considered to interfere. If the samples showed significant levels of interference, the results were reviewed prior to proceeding. PPV titers measured as PFU/mL were reduced by  $> 5$  logs for both Fresh Frozen Plasma and Platelets as shown in chart #5.

#### Fresh Frozen Plasma

Three units of fresh frozen plasma (FFP, approximately 300 mL / unit) were pooled. The pooled mixture was then spiked with 1% of PPV stock solution (exact volumes were recorded at the time of spiking). The spiked starting materials were then divided into three, 100 mL samples and ten, 50 mL samples.

A 6 mL sample of the spiked starting material was removed from one of the 100 mL aliquots, adjusted to pH 6.5 - 7.5 (if necessary) and divided into multiple aliquots. One aliquot was tested immediately, the remaining aliquots were snap frozen, and stored as backups at or below  $-70^{\circ}\text{C}$ . The remaining 94 mL of sample was held at ambient temperature for the duration of the process. Following incubation, a 6 mL sample was removed, adjusted to pH 6.5 - 7.5 (if necessary) and divided into multiple aliquots. One aliquot was tested immediately, the remaining aliquots were snap frozen, and stored as backups at or below  $-70^{\circ}\text{C}$ . The remaining spiked samples were then treated with 282 nm monochromatic light in the apparatus of Figs. 1-4.

Following treatment, a 6 mL aliquot was removed from each treated sample, adjusted to pH 6.5 - 7.5 (if necessary) and divided into multiple aliquots. One

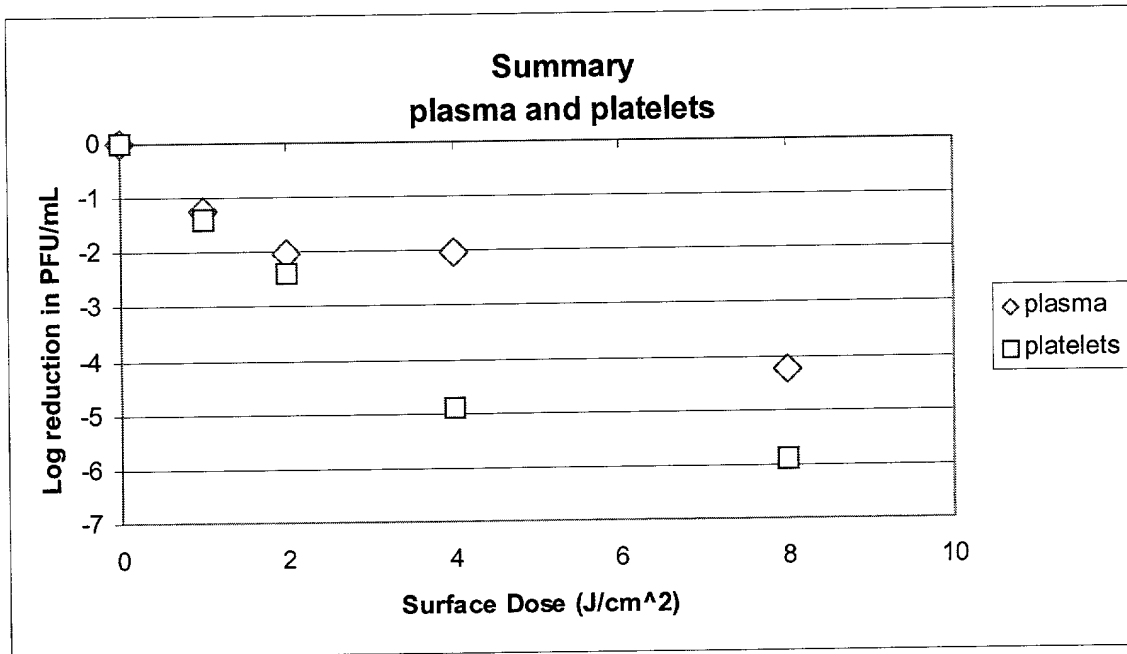
aliquot was tested immediately, the remaining aliquots were snap frozen and stored as backups at or below -70°C.

#### Platelets, RPC

Six units of random platelet concentrates (RPC, approximately 65 mL / unit) were pooled. The pooled mixture was then spiked with 1% of PPV stock solution (exact volumes were recorded at the time of spiking). The spiked material was then divided into three, 60 mL samples and seven, 20 mL samples. A 6 mL sample of the spiked starting material was removed from one of the 60 mL aliquots, adjusted to pH 6.5 - 7.5 (if necessary) and divided into multiple aliquots. One aliquot was tested immediately, the remaining aliquots were snap frozen, and stored as backups at or below -70°C. The remaining 54 mL of sample was held at ambient temperature for the duration of the process. Following incubation, a 6 mL sample was removed, adjusted to pH 6.5 - 7.5 (if necessary) and divided into multiple aliquots. One aliquot was tested immediately, the remaining aliquots were snap frozen, and stored as backups at or below -70°C. The remaining spiked samples were then treated with 282 nm monochromatic light in the apparatus of Figs. 1-4.

Following treatment, a 6 mL aliquot was removed from each treated sample, adjusted to pH 6.5 - 7.5 (if necessary) and divided into multiple aliquots. One aliquot was tested immediately, the remaining aliquots were snap frozen, and stored as backups at or below -70°C.

## Results



For both platelets and plasma, the PPV was reduced to non-detectable levels. The maximum log removal shown is that which could be quantified given the limits of detection. Extra wells were set for the highest dose measurements, and no virus was detected.

Test results are reflected in the following Tables.

TABLE 2  
Log<sub>10</sub> Reduction Summary - Fresh Frozen Plasma

Virus	Sample Designation	Log <sub>10</sub> Reduction
PPV	FFP-Treatment A (100 mL)	2.24 ± 0.18
	FFP-Treatment B (100 mL)	3.14 ± 0.16
	FFP-Treatment C (50 mL)	1.27 ± 0.13
	FFP-Treatment D (50 mL)	0.97 ± 0.27
	FFP-Treatment E (50 mL)	2.03 ± 0.24
	FFP-Treatment F (50 mL)	1.02 ± 0.18
	FFP-Treatment G (50 mL)	2.04 ± 0.25
	FFP-Treatment H (50 mL)	1.34 ± 0.13
	FFP-Treatment I (50 mL)	4.24 ± 0.25
	FFP-Treatment J (50 mL)	2.47 ± 0.22
	FFP-Treatment K (50 mL)	* > 4.87 ± 0.12
	FFP-Treatment L (50 mL)	* > 4.87 ± 0.12

5      \* Virus reduced to non-detectable levels.

TABLE 3  
Log<sub>10</sub> Reduction Summary - Random Platelet Concentrates

Virus	Sample Designation	Log <sub>10</sub> Reduction
PPV	RPC-Treatment A (60 mL)	2.35 ± 0.21
	RPC-Treatment B (60 mL)	4.14 ± 0.21
	RPC-Treatment C (20 mL)	1.43 ± 0.31
	RPC-Treatment D (20 mL)	2.42 ± 0.36
	RPC-Treatment E (20 mL)	4.90 ± 0.19
	RPC-Treatment F (20 mL)	2.95 ± 0.19
	RPC-Treatment G (20 mL)	* > 5.93 ± 0.16
	RPC-Treatment H (20 mL)	4.45 ± 0.19
	RPC-Treatment I (20 mL)	* > 5.93 ± 0.16

10      \* Virus reduced to non-detectable levels.

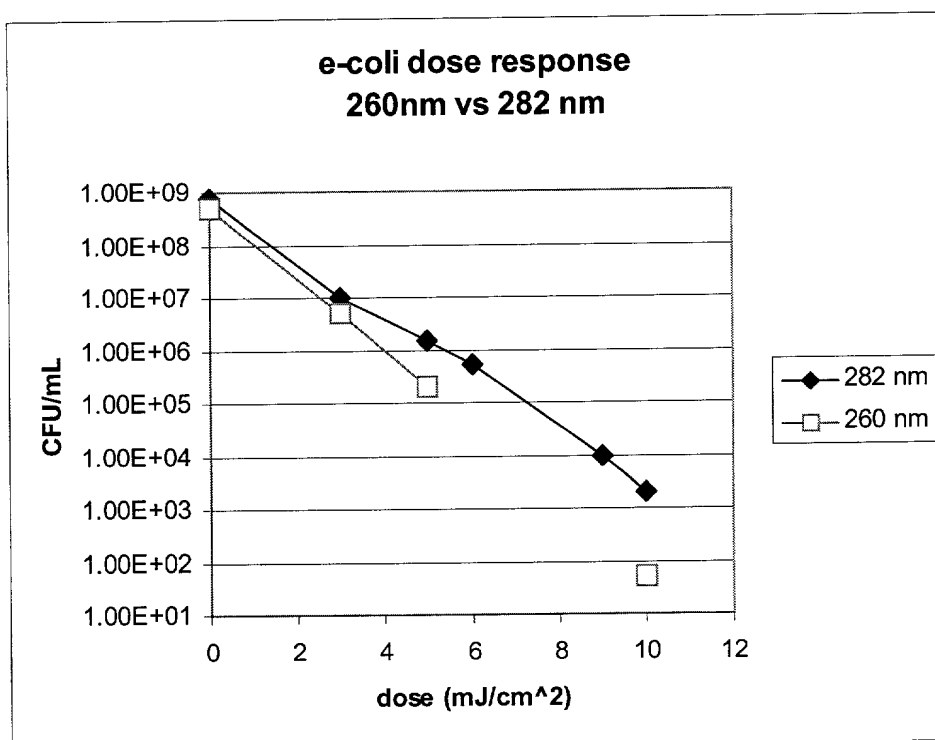
Based on the foregoing test results, it is clear that substantially monochromatic light at a wavelength of 282 nm delivered in the disclosed treatment system is effective against refractory virus in both plasma and platelets.

### EXAMPLE 3

In order to establish the effectiveness of longer wavelength light produced by the XeBr source, tests were made comparing inactivation rates at 282 nm to those measured at 260 nm. 260 nm is near the peak of the germicidal action spectra.

- 5 Monochromatic light is produced using an excimer lamp using  $\text{Cl}_2$  to produce light at comparable intensities at 260 nm. A lab cultured coli was used.

A collimated beam apparatus was used to deliver light to static sample containers. Heterotrophic plates counts (HPC) were performed as per Standard Methods Spread Plate Method (9215 C.) (19<sup>th</sup> Ed). This included serial diluting the sample by 10s  
10 using 50% Difco nutrient broth as diluent, pipetting 100  $\mu\text{L}$  of each dilution onto the plates, and spreading the sample with a plastic disposable spreader so that the entire sample is absorbed into the agar media. Samples were incubated (plates inverted) at 32-35°C and counted for quantification of heterotrophic bacteria at 24 hours and again at 48 hours and recorded. Only the analytical plate (that with colonies between 30 and 300)



was reported although all counts were recorded in the lab book. The resulting data, which reflects averages of several runs, is shown in the plot of Figure 7.

\* \* \* \* \*

Having thus described preferred embodiments and exemplary

5 uses/applications of the present disclosure, it is to be understood that the specifically disclosed forms are merely illustrative of the scope of the present disclosure. Various changes may be made in the function and arrangement of parts and processing parameters; equivalent means may be substituted for those described and/or illustrated; and certain features may be used independently from others without departing from the  
10 spirit and scope of the invention as defined in the claims that follow.